



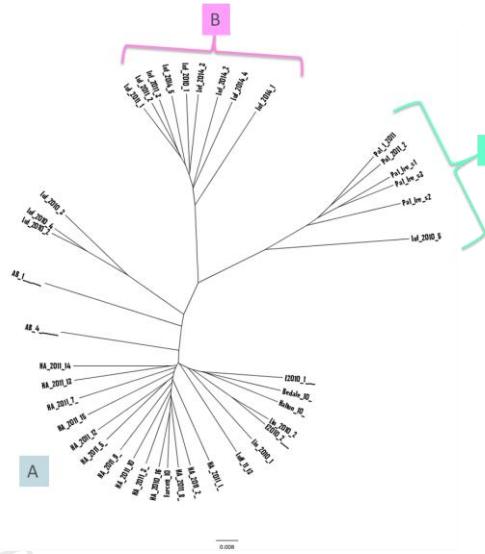
Ulrike Gartner

Flash-talk Corvallis 12 March 2019



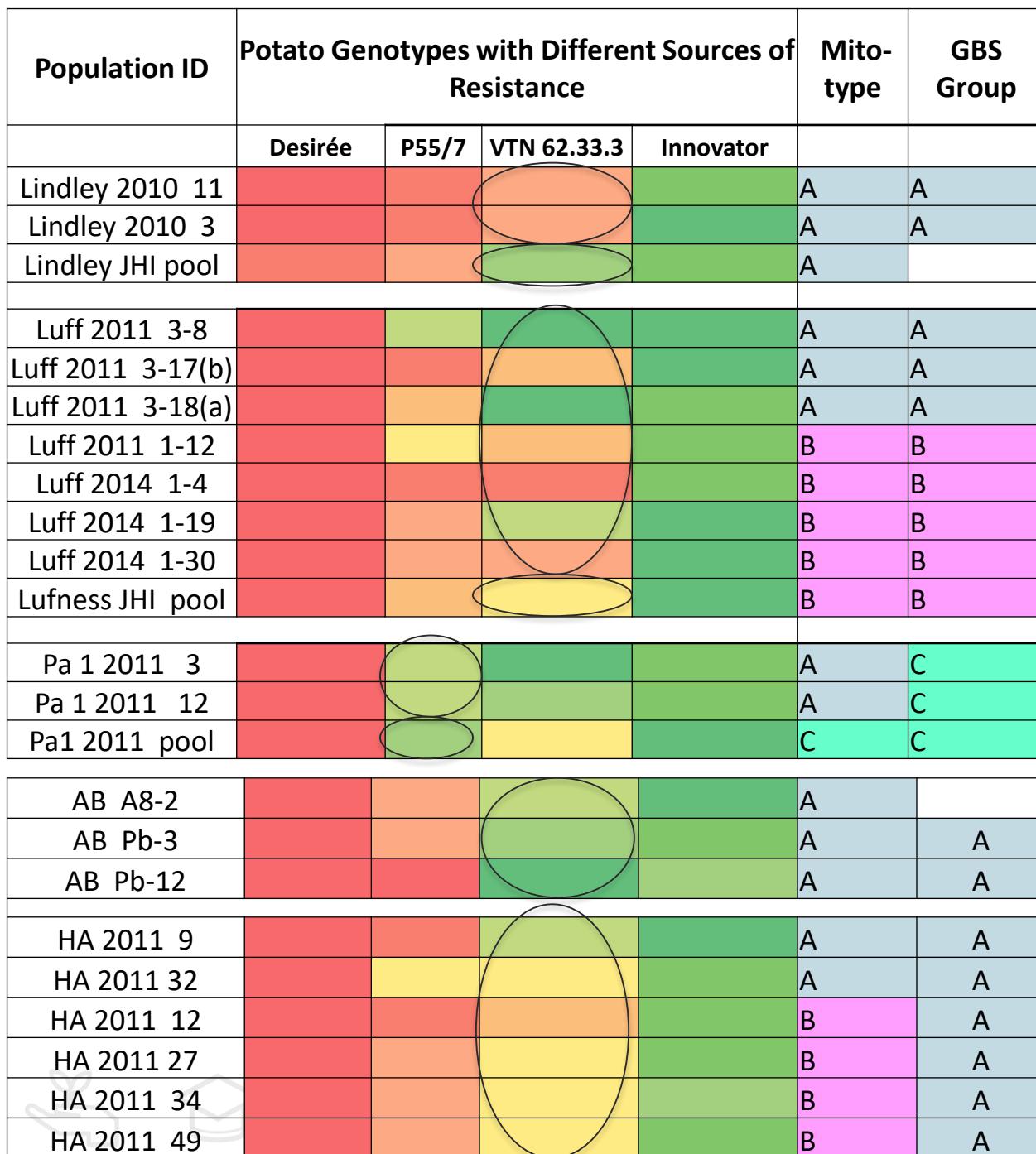
Can mitochondrial markers that distinguish the different introductions be used as pathotype markers?

Samples	Population	Type A	Type B	Type C	No. cysts analysed
JHI collection	Lindley	25 (96%)	1 (4%)		26
	Luffness field 1	2 (6%)	33 (94%)		35
	Pa1	4 (19%)	2 (10%)	15 (71%)	21
Field	Luffness field 1	7 (39%)	9 (50%)	2 (11%)	18





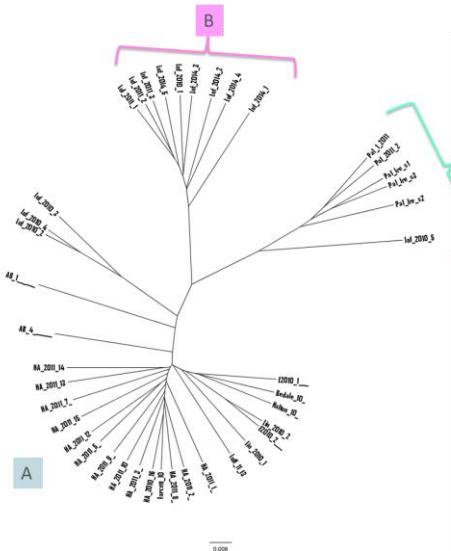
The James
Hutton
Institute



Level of
Resistance



Future Work: Development of molecular pathotype markers for *G. pallida*



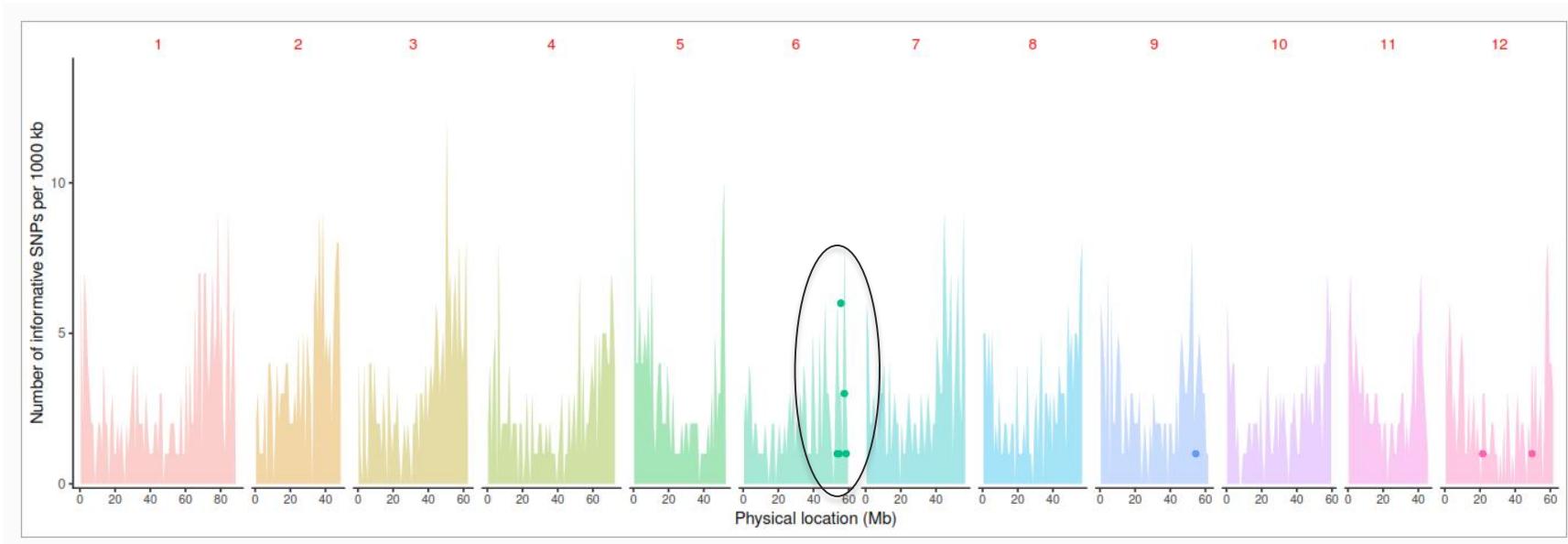
Single cyst line/population	Pheno type	Mito type	GBS group
HA 12	PA3	B	A
HA27	PA3	B	A
HA34	PA3	B	A
PB-12	PA2	A	A
Luff3-18	PA2	A	A
Luff 3-8	PA2	A	A
Luff1-12	PA3	B	B
Luff1-4	PA3	B	B
Luff1-30	PA3	B	B
Pa1-12	PA1	A	C
Pa1 pool	PA1	C	C
Pa1 ire 1	PA1	n/a	C

	Allele1/2	% allele 1 in populations		
		PA2	PA3	PA1
1	A/T	0.0	0.0	100.0
2	T/C	0.0	9.1	100.0
3	G/A	100.0	100.0	16.7
4	G/A	100.0	100.0	16.7
5	G/A	100.0	100.0	16.7
6	T/C	0.0	9.1	100.0
7	G/C	87.5	81.8	16.7
8	T/C	100.0	9.1	100.0
9	T/C	25.0	81.8	0.0
10	T/A	37.5	90.9	0.0
11	T/C	50.0	81.8	0.0
12	G/A	75.0	90.9	16.7



The diploid *S. spegazzinii*, acc. 7195, shows natural resistance to *G. pallida*.

Generic-mapping enrichment Sequencing (GenSeq)

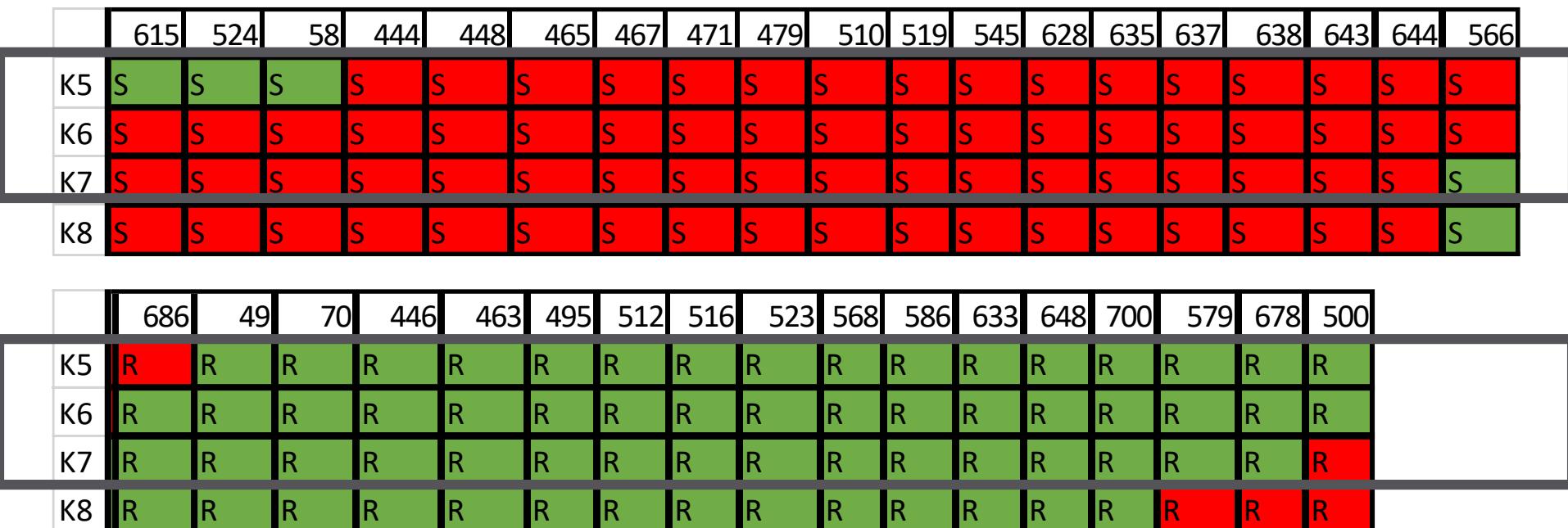


Graphical Genotyping

	SP	RP
K5	S	R
K6	S	R
K7	S	R
K8	S	R

K5 to K8 comprise 5.4 Mb

K5 to K7 comprise 3.4 Mb



S	susceptible
R	resistant
RR	homozygous resistant
SR	heterozygous resistant



Thank you

